



OPEN

Behavioral characteristics of dopamine D₅ receptor knockout mice

Hitomi Sasamori^{1,7}, Toshiaki Asakura^{2,7}, Chiaki Sugiura^{1,7}, Youcef Bouchekioua¹, Naoya Nishitani^{1,3}, Masaaki Sato¹, Takayuki Yoshida^{1,4}, Miwako Yamasaki⁵, Akira Terao⁶, Masahiko Watanabe⁵, Yu Ohmura^{1✉} & Mitsuhiro Yoshioka^{1,8}

Major psychiatric disorders such as attention-deficit/hyperactivity disorder and schizophrenia are often accompanied by elevated impulsivity. However, anti-impulsive drug treatments are still limited. To explore a novel molecular target, we examined the role of dopamine D₅ receptors in impulse control using mice that completely lack D₅ receptors (D5KO mice). We also measured spontaneous activity and learning/memory ability because these deficits could confound the assessment of impulsivity. We found small but significant effects of D₅ receptor knockout on home cage activity only at specific times of the day. In addition, an analysis using the q-learning model revealed that D5KO mice displayed lower behavioral adjustment after impulsive actions. However, our results also showed that baseline impulsive actions and the effects of an anti-impulsive drug in D5KO mice were comparable to those in wild-type littermates. Moreover, unlike previous studies that used other D₅ receptor-deficient mouse lines, we did not observe reductions in locomotor activity, working memory deficits, or severe learning deficits in our line of D5KO mice. These findings demonstrate that D₅ receptors are dispensable for impulse control. Our results also indicate that time series analysis and detailed analysis of the learning process are necessary to clarify the behavioral functions of D₅ receptors.

Various psychiatric disorders, such as attention-deficit/hyperactivity disorder (ADHD), schizophrenia, substance use disorder, bipolar disorder, and borderline personality disorder, have been associated with increased impulsivity¹. Clinically available anti-impulsive drugs vary from country to country, but at present, the major ones include amphetamine, methylphenidate, and lisdexamfetamine as psychostimulants, and atomoxetine, guanfacine, and clonidine as adrenaline-related drugs. However, psychostimulants pose a risk of abuse and dependence, and adrenaline-related drugs are often difficult to recommend due to their cardiovascular and autonomic side effects. For example, atomoxetine, a noradrenaline reuptake inhibitor, might not be appropriate in some patients since it can exacerbate hypertension, which is often comorbid with ADHD, bipolar disorder, and borderline personality disorder^{2–4}. Although guanfacine and clonidine are hypotensive agents, their sedative side effects could interfere with work or study. Therefore, further development of novel anti-impulsive drugs is required.

To this end, we examined whether dopamine D₅ receptors play an important role in the control of impulsivity. We have previously demonstrated that dopamine D₁-like receptors in the ventral part of the medial prefrontal cortex play a critical role in the anti-impulsive effects of milnacipran, duloxetine, and atomoxetine^{5,6}. There are two types of dopamine D₁-like receptors: D₁ receptors and D₅ receptors. Dopamine D₁ receptors are densely expressed in the nucleus accumbens, where impulsivity is enhanced by increased extracellular dopamine levels^{7,8}, while dopamine D₅ receptors are sparsely expressed in the region⁹. In comparison, in the medial prefrontal cortex, where impulsivity is inhibited by increased extracellular dopamine levels^{10,11}, both dopamine D₁ and dopamine D₅ receptors are expressed⁹. Furthermore, dopamine D₅ receptors have a tenfold higher affinity for dopamine

¹Department of Neuropharmacology, Hokkaido University Faculty of Medicine, Sapporo, Japan. ²Hokkaido University School of Medicine, Sapporo, Japan. ³Laboratory of Molecular Pharmacology, Institute of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Kanazawa, Japan. ⁴Department of Neurophysiology, Graduate School of Biomedical and Health Sciences, Hiroshima University, Hiroshima, Japan. ⁵Department of Anatomy, Hokkaido University Faculty of Medicine, Sapporo, Japan. ⁶Department of Biology, School of Biological Sciences, Tokai University, Sapporo, Japan. ⁷These authors contributed equally: Hitomi Sasamori, Toshiaki Asakura, and Chiaki Sugiura. ⁸Mitsuhiro Yoshioka is deceased. ✉email: yohmura@med.hokudai.ac.jp

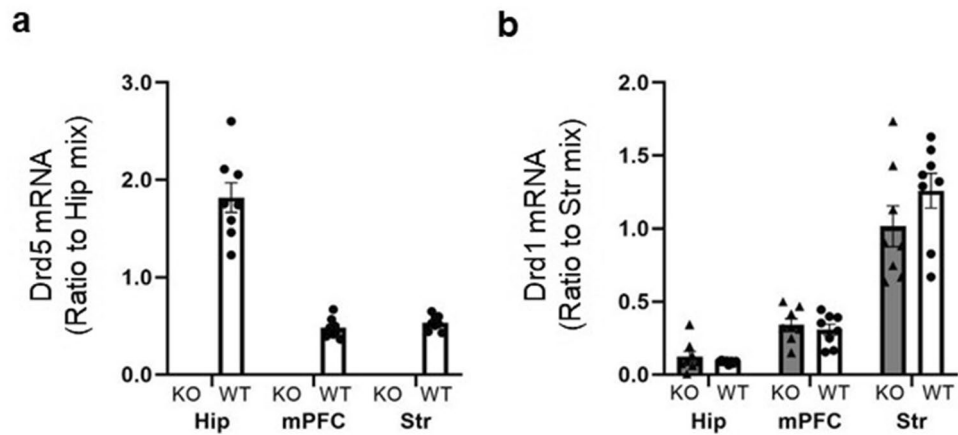


Figure 1. Effects of genotype on *Drd5* and *Drd1* gene expression. **(a)** *Drd5* gene relative expression levels in D5KO (KO, dark bars) mice and wildtype (WT, white bars) littermates. **(b)** *Drd1* gene relative expression levels in D5KO (KO, dark bars) mice and WT (white bars) littermates. Mix means a mixture of KO and wildtype samples. Hip: hippocampus, mPFC: medial prefrontal cortex, Str: striatum. The data are presented as the means \pm SEM.

than dopamine D₁ receptors¹². Thus, we hypothesized that the anti-impulsive effects of the above drugs might be exerted via stimulation of dopamine D₅ receptors.

The development of a selective dopamine D₅ receptor agonist might resolve problems encountered with the current stable of anti-impulsive drugs by enabling us to selectively manipulate the medial prefrontal cortex without affecting the nucleus accumbens. To our knowledge, however, there are so far no drugs that clearly distinguish between dopamine D₁ and D₅ receptors. Given that psychostimulants primarily facilitate addiction through the modulation of the nucleus accumbens¹³, a selective dopamine D₅ receptor agonist might not induce this process, unlike psychostimulant-based anti-impulsive drugs. Furthermore, a selective agonist for dopamine D₅ receptors would not likely exacerbate hypertension since dopamine D₅ receptor knockout (D5KO) mice are hypertensive¹⁴. Spontaneous motor activity in D5KO mice is generally normal or reduced, implying that a selective dopamine D₅ receptor agonist will not induce sedation. However, in the absence of selective D₅ receptor agonists, examining dopamine D5KO mice is a reasonable way to determine whether D₅ receptors could be a promising target for anti-impulsive drugs.

In the present study, we used an alternative line of D5KO mice¹⁵ instead of traditional D5KO mice¹⁴ because of three reasons. First, the traditional D5KO mice could express truncated transcripts that might alter the expression of related genes^{16,17}, while the alternative line of D5KO mice would not express them because the entire dopamine D₅ receptor gene region, including the promoter region, is removed. Second, previous studies have shown that different lines of transgenic mice or different background strains of transgenic mice could alter baseline behavioral phenotype^{18,19}. To clarify the role of a molecule in brain functions, we are better off testing not only a specific line or background strain but also another line or background strain. Third, some studies have reported lower spontaneous motor activity and deficits of learning and working memory in traditional D5KO mice^{20–22}. These phenotypes make it difficult for researchers to assess impulsivity because most tasks evaluating impulsivity assume a certain level of spontaneous activity and learning/memory ability. We speculated that these phenotypes are due to the above reasons, but not due to the lack of D₅ receptors.

In this study, using an alternative line of D5KO mice, we conducted quantitative PCR to confirm that dopamine D₅ receptors were not transcribed as expected and whether compensatory changes in dopamine D₁ receptors did not occur, (2) measured locomotor activity in two different environments: a novel environment and a familiar environment, (3) conducted a Y-maze test to assess working memory, and (4) employed the 3-choice serial reaction time task (3-CSRTT)^{11,23} to assess learning ability and impulsivity. To evaluate possible learning deficits or bias, we modeled the learning process within the 3-CSRTT using a q-learning model.

Results

RNA analysis. To confirm that the dopamine D₅ receptor gene is not transcribed and that a compensatory overexpression of D₁ receptors does not occur, we conducted quantitative PCR tests. As expected, the *Drd5* gene expression levels were below the detection limit in the D5KO mice (Fig. 1a). Moreover, the *Drd1* gene expression levels were not increased in the D5KO mice compared to wildtype littermates in the hippocampus ($t_{14}=0.96$, $p=0.35$), medial prefrontal cortex (mPFC) ($t_{14}=0.61$, $p=0.55$), and striatum ($t_{14}=-1.34$, $p=0.20$) (Fig. 1b).

Home cage activity. To measure locomotor activity in a familiar environment, we measured home cage activity for 24 h. We performed a three-factor ANOVA on the changes in locomotor activity every two hours in their home cages (Fig. 2a). There was a main effect of time ($F_{5,39, 285.43}=115.28$, $p<0.001$, with Greenhouse–Geisser correction). There was a significant interaction between time and genotype ($F_{1, 53}=3.34$, $p<0.001$, with Greenhouse–Geisser correction). Other main effects and interactions were not detected (Table S1).

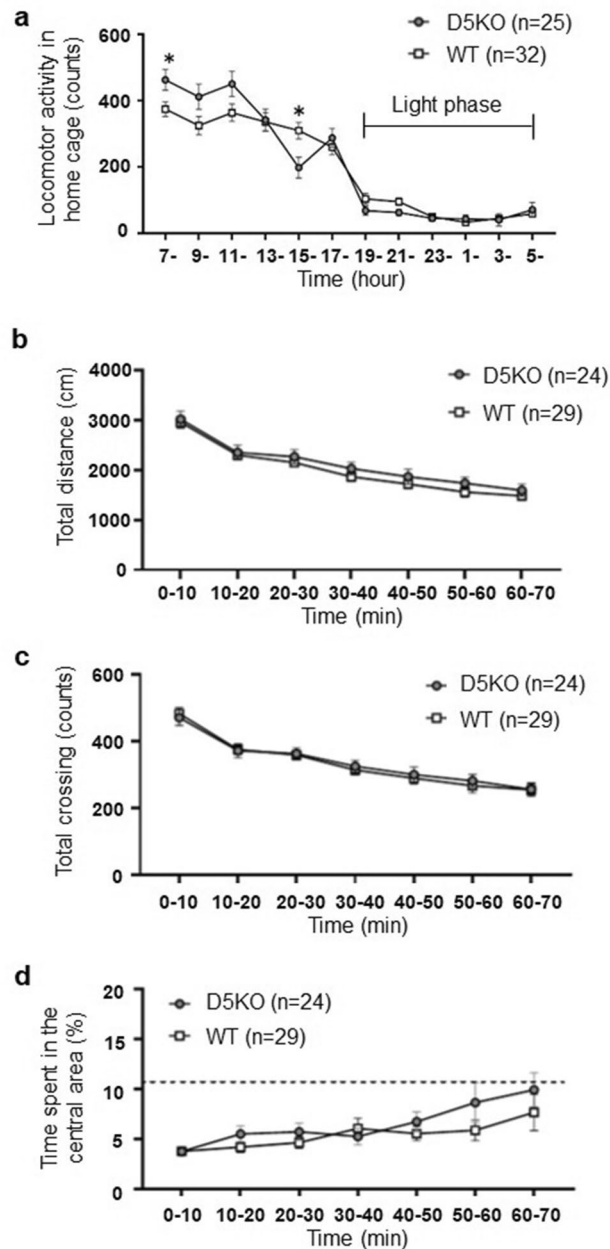


Figure 2. Effects of genotype on 24 h locomotor activity in home cages and on parameters in the open field test. (a) Home cage locomotor activity of D5KO mice and wildtype littermates every 2 h. * $p < 0.05$. (b) The total distance traveled in the open field test was divided into seven time phases (10 min bins). (c) The number of total crossings (crossings of the lines made by the division of the field [45 × 45 cm] into 7.5 cm × 7.5 cm squares) in the open field test was divided into seven time phases (10 min bins). (d) The percentage of time spent in the central area, a measure of decreased anxiety-like behavior in the open field test, was divided into seven time phases (10 min bins). The filled circles indicate D5KO mice and white squares indicate WT littermates. The data are presented as the means ± SEM.

Simple main effects analyses for each time point revealed that D5KO mice were significantly more active than wildtype littermates between 7:00 and 9:00 ($F_{1,53} = 4.63$, $p = 0.036$) (Fig. 2a). D5KO mice were also significantly less active than wildtype littermates between 15:00–17:00 ($F_{1,53} = 6.88$, $p = 0.011$) (Fig. 2a). No differences by genotype at other times were detected ($F_{s1,53} < 3.19$, $p > 0.07$).

Open field test. To measure locomotor activity in a novel environment, we conducted open field tests for 70 min. Three factor ANOVA revealed that the distance traveled over the testing period significantly decreased

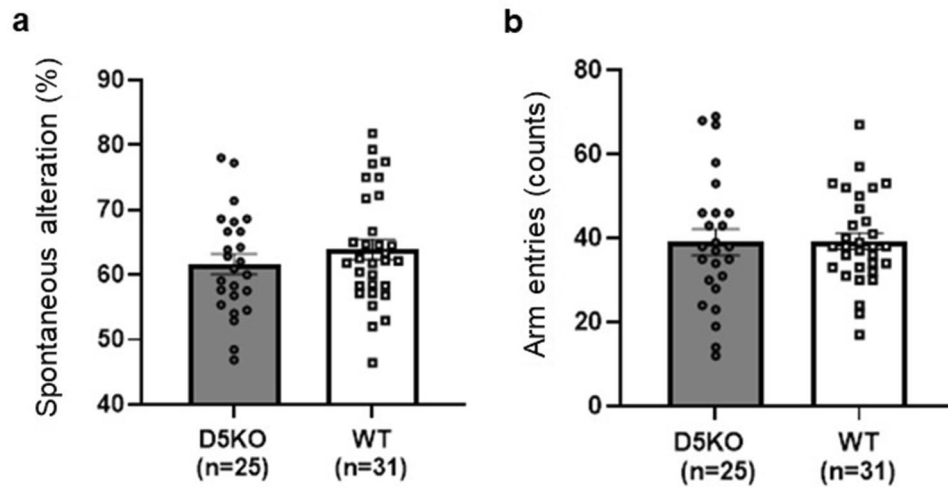


Figure 3. The effects of dopamine D₅ receptor KO on the parameters in the Y maze test. **(a)** The percentage of spontaneous alternation, a measure of working memory, in D5KO mice and their WT littermates. **(b)** The total number of arm entries, a measure of locomotor activity, of D5KO mice and their WT littermates. The data are presented as the means ± SEM.

over time ($F_{3,45,169} = 119.1$, $p < 0.0001$, with Geisser-Greenhouse correction). However, we found no significant main effects or interactions in other factors (Fig. 2b, Table S2).

A three-factor ANOVA for the number of crossings revealed a significant main effect of time ($F_{6,294} = 93.96$, $p < 0.0001$). However, we found no significant main effects or interactions in other factors (Fig. 2c, Table S2).

A three-factor ANOVA for the percentage of time spent in the central area, which is a measure of decreased anxiety-like behavior, revealed a significant main effect of time ($F_{2,99,146.5} = 6.86$, $p = 0.0002$, with Geisser-Greenhouse correction). However, we did not find other main effects or any interactions (Fig. 2d, Table S2).

Y maze test. To assess working memory in mice, we conducted the Y maze test. Two factor ANOVA for the percentage of spontaneous alternation, a measure of working memory, did not reveal any main effects or interaction (Fig. 3a, Table S3). Two factor ANOVA for the total number of arm entries, a measure of locomotor activity, did not reveal any main effects or interaction (Fig. 3b, Table S3).

Assessment of learning ability with a q-learning model. To determine whether D5KO mice display learning deficits or bias, we modeled the learning process of 3-CSRTT. We assume experience and non-reward distributions to represent premature behaviors of mice. Experience distribution represents a memory state of all trials and is updated by the q-learning process whatever a previous result is, while non-reward distribution represents a memory state of premature responses. Table 1 shows the estimated parameters of a q-learning model, and Table S4 has descriptions of each parameter. In this model, key parameters are learning rates, representing learning ability, and an inverse temperature, representing confidence in their own choice. The baseline effect of learning rate for the experience distribution, $\alpha_{X,0}$, was 0.04296 while that for the non-reward distribution, $\alpha_{Y,0}$, was 0.08555. For the experience distribution, only male effects were significant according to the 95% Highest Density Interval (HDI). Also, for the non-reward distribution, both D5KO and male effects were negative and significant, based on 95% HDI. This result indicated that D5KO mice have a learning deficit for premature results, not for non-premature results. As for the inverse temperature, baseline effect, β_0 , took a value of 130.603, and both D5KO and male effects were not significant.

Although $\alpha_{X,male}$, $\alpha_{Y,D5KO}$, and $\alpha_{Y,male}$ were not zero for these 95% HDI, the degree of effects of these parameters remains unclear. To quantify contributions of these parameters, we simulated the q-learning process with estimated parameters. Table S5 shows proportions of premature responses for each session for trial and simulation data. For the simulation data, overall, the start timing of the proportions was consistently lower than those for trial data. The proportions of premature responses among each session fluctuated for real and simulated results. If we focus on the values at session 10, the minimum values were observed in D5KO male mice, whereas the maximum value was observed in wildtype female mice. Simulated results with individually estimated parameters using the q-learning model over trials indicated the model could potentially capture behaviors of trial results in the 3-CSRTT (see Supplementary Fig. 1).

Averaged values for functions used in the q-learning model at the end of the simulation with estimated parameters are shown in Fig. 4. Values at each elapsed time were averaged values calculated from 100 simulation results. Overall, there were no major differences in the shape of the functions. For the probability of confidence, the rise of the distribution around 5 s was steeper in D5KO mice than in wild type mice, reflecting the higher value of inverse temperature, although the D5KO effect for an inverse temperature is not significant.

	Mean	95% HDI ^a
$\alpha_{X,0}$	0.04296	(0.03829, 0.04784)
$\alpha_{X,D5KO}$	- 0.00147	(- 0.00379, 0.00058)
$\alpha_{X,male}$	- 0.00612	(- 0.00895, - 0.00372)
$\alpha_{Y,0}$	0.08555	(0.07465, 0.09726)
$\alpha_{Y,D5KO}$	- 0.00517	(- 0.00782, - 0.00260)
$\alpha_{Y,male}$	- 0.01394	(- 0.01726, - 0.01087)
β_0	130.603	(117.447, 144.407)
β_{D5KO}	9.925	(- 2.731, 22.702)
β_{male}	- 5.961	(- 20.987, 8.380)
α_M	0.03755	(0.03097, 0.04463)
σ_M^2	617.027	(526.455, 718.198)
σ_r^2	433.462	(373.520, 502.798)
$\sigma_{X,0}^2$	15.816	(12.792, 19.261)
$\sigma_{Y,0}^2$	52.144	(43.677, 62.984)

Table 1. Estimated parameters of the q-learning model. It is noted that $\alpha_{X,D5KO}$, $\alpha_{X,male}$, $\alpha_{Y,D5KO}$, and $\alpha_{Y,male}$ were sampled from real numbers, while the other parameters were sampled from positive real numbers. ^aHDI represents the highest density interval.

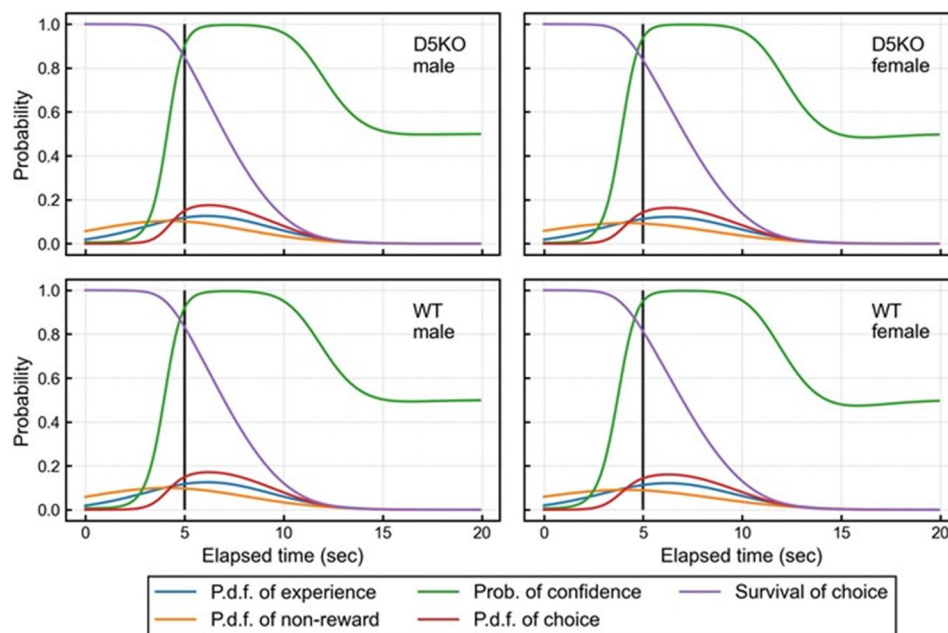


Figure 4. Averaged function values for the q-learning model at the end of the simulation with estimated parameters. Each simulation was run for 900 steps and 100 simulations were performed. Figure contains averaged values for these simulations at each elapsed time (s). The black line represents the flash timing of the trial. Each line represents the following: Probability density function (p.d.f.) of experience (blue); the p.d.f. of experience distribution. P.d.f. of non-reward (orange); the p.d.f. of non-reward distribution. Prob. of confidence (green); the probability of confidence at each elapsed time. P.d.f. of choice (red); the p.d.f. of choice distribution. Survival of choice (purple): the survival function of the choice distribution.

Effects of acute duloxetine administration on impulsive action. To assess the effects of D5KO on impulsive action and anti-impulsive effects of duloxetine, we administered duloxetine and conducted the 3-CSRTT. Duloxetine administration reduced the percentage of premature responses in a dose-dependent manner (Fig. 5a,b). Three factor repeated measures ANOVA revealed a significant dose effect on the percentage of premature responses (Fig. 5a,b) ($F_{2,51,140,26} = 14.06$, $p < 0.001$, with Greenhouse–Geisser correction). A multiple comparison with Bonferroni's correction revealed that the 0.3 mg/kg, 1.0 mg/kg, and 3.0 mg/kg dose of duloxetine significantly decreased the percentage of premature responses compared to vehicle administration. However, we did not find any other main effects or interactions (Table S6).

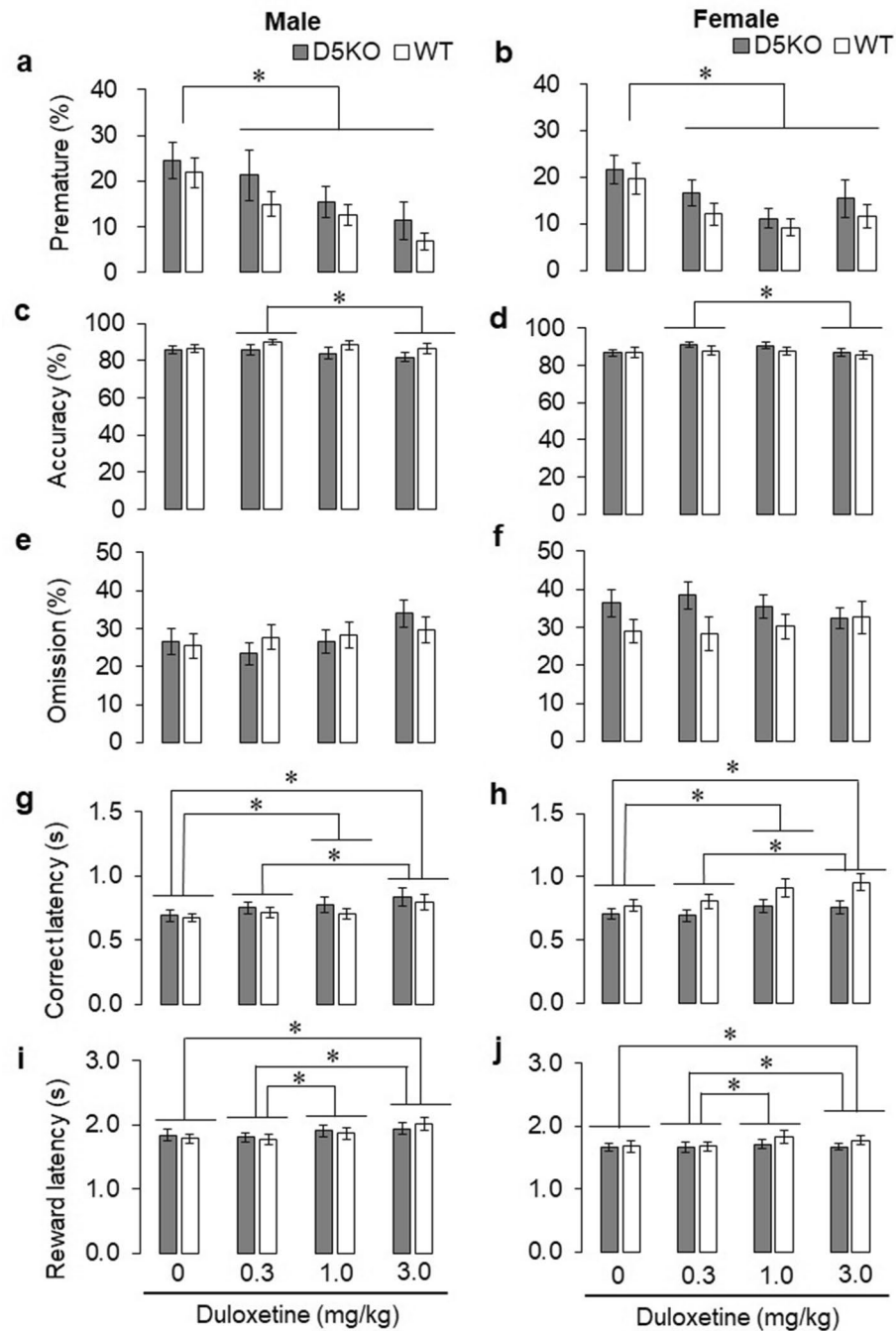


Figure 5. Effects of acute duloxetine administration on impulsive action. **(a)** The percentage of premature responses, a measure of impulsive action, of male D5KO mice and their WT littermates. **(b)** The percentage of premature responses, a measure of impulsive action, of female D5KO mice and their WT littermates. **(c)** Accuracy, the percentage of correct responses of male D5KO mice and their WT littermates. **(d)** Accuracy, the percentage of correct responses of female D5KO mice and their WT littermates. **(e)** The percentage of omissions of male D5KO mice and their WT littermates. **(f)** The percentage of omissions of female D5KO mice and their WT littermates. **(g)** The mean correct response latency (s) of male D5KO mice and their WT littermates. **(h)** The mean correct response latency (s) of female D5KO mice and their WT littermates. **(i)** The mean reward response latency (s) of male D5KO mice and their WT littermates. **(j)** The mean reward response latency (s) of female D5KO mice and their WT littermates. The black bars indicate D5KO mice and white bars indicate WT littermates. The data are presented as the means \pm SEM. * $p < 0.05$.

Furthermore, other parameters were also affected by the administration of duloxetine. Three factor repeated measures ANOVA revealed a significant dose effect on accuracy (Fig. 5c,d) ($F_{2,56, 143,28} = 3.93, p = 0.014$, with Greenhouse–Geisser correction), correct response latency (Fig. 5g,h) ($F_{2,40, 134,19} = 10.66, p < 0.001$, with Greenhouse–Geisser correction), and reward latency (Fig. 5i,j) ($F_{3,168} = 6.74, p < 0.001$), but not on omission (Fig. 5e,f). Multiple comparisons with Bonferroni's correction revealed that the 3.0 mg/kg dose of duloxetine decreased accuracy compared to the 0.3 mg/kg dose (Fig. 5c,d), while the 1.0 mg/kg dose of duloxetine prolonged correct response latency compared to vehicle, the 3.0 mg/kg dose of duloxetine prolonged correct response latency compared to vehicle and the 0.3 mg/kg dose (Fig. 5g,h), the 3.0 mg/kg dose of duloxetine administration significantly prolonged reward latency compared to vehicle and 0.3 mg/kg duloxetine, and the 1 mg/kg duloxetine administration significantly prolonged the reward latency compared to 0.3 mg/kg (Fig. 5i,j). Moreover, a main effect of sex was detected on mean reward response latency ($F_{1,56} = 5.210, p = 0.026$). However, we did not find any other main effects or interactions (Table S6).

Discussion

Although minor differences were found, no major differences were observed in any behavioral parameters between wildtype and D5KO mice. Small but significant effects of D5KO were observed in home cage activity only at specific times of day. In addition, D5KO mice displayed lower behavioral adjustments after premature responses in the 3-CSRTT. We did not observe a reduction in locomotor activity in a novel environment or working memory deficits in D5KO mice, inconsistent with some previous studies. No significant effects of D5KO on impulsive action were observed, suggesting that our hypothesis that D₅ receptors play an essential role in impulse control is incorrect. We discuss possible interpretations for each result below.

We replicated a previous KO study that indicated no mRNA expression of D₅ receptors using Northern blotting¹⁵. We used a different method, quantitative RT-PCR (Fig. 1a) and reached the same conclusion: our D₅ receptor KO mice do not express D₅ receptor at all. Another concern of D₅ receptor KO mice is compensatory effects. For decades, studies from transgenic and gene knockout mice have contributed to the delineation of the functional role of many kinds of proteins. However, recent evidence has demonstrated that the interpretation of these studies may be complicated by compensatory changes in animals because gene mutations truncating the encoded protein could affect the expression of related genes^{16,17}. Our RT-qPCR results (Fig. 1b) indicated that we could exclude the possibility of a compensatory increase of dopamine D₁ receptors, which are involved in impulse control. However, we cannot deny the other numerous possibilities that expression of other genes²⁴ or functional pathways^{25,26} was altered in D₅ receptor KO mice. In future studies, an AAV-mediated knockdown or knockout of the D₅ receptor in the PFC would be required to ensure no compensatory changes because testing above numerous possibilities is impractical.

We found that dopamine D₅ receptor KO mice exhibited higher locomotor activity at the beginning of the dark period (7:00–9:00) but lower locomotor activity at the end of the dark period (15:00–17:00) in their home cage, a familiar environment (Fig. 2a). Previous studies have shown that dopamine D₅ receptor KO mice display lower locomotor activity than wildtype mice^{21,22}, while others did not detect any difference in locomotor activity^{15,20}. The present study might explain the inconsistent results from previous studies on locomotor activity in dopamine D₅ receptor KO mice. In the previous studies, an open field test lasting 60 to 150 min has been used to measure locomotor activity. However, since the locomotor activity of dopamine D₅ receptor KO mice changed significantly between the first and second halves of the dark period, the results may vary depending on the time of day the test was conducted. Although speculative, previous studies that showed lower locomotor activity in dopamine D₅ receptor KO mice might have been conducted in the latter half of the dark period.

In the open field test in the present study, we did not detect any difference in locomotor activity in the novel environment between dopamine D5KO mice and their wild type littermates (Fig. 2b,c). This might be due to the fact that the time of measurement was not kept constant. Alternatively, the results in the home cage described above might be limited to a familiar environment. Because further studies examining locomotor activity in the open field at specific times will be required to address this issue, we suspend our conclusion. In addition, there was no difference in the time spent in the central compartment (%), a measure of reduced anxiety-like behavior, in dopamine D₅ receptor KO mice compared to their wildtype littermates (Fig. 2d). Therefore, our findings indicate that dopamine D₅ receptors may not relate to anxiety-like behavior, consistent with previous studies^{21,22}.

In the Y maze test, there was no difference in working memory in dopamine D₅ receptor KO mice compared to their wildtype littermates (Fig. 3). However, in previous studies, dopamine D₅ receptor KO mice tended to exhibit lower working memory^{20–22}. There are at least two possible explanations for this discrepancy. First, we used an alternative line of D₅ receptor KO mice in this study¹⁵, while the previous studies that detected working memory deficits had used traditional dopamine D₅ receptor KO mice¹⁴. As discussed earlier, the traditional mice could alter the expression of related genes^{16,17}. Thus, working memory deficits observed in these studies might be due to the secondary effects. The second possibility is the difference in the working memory measurement task employed. A previous study demonstrating working memory deficit in D₅ receptor KO mice used a baited T-maze test²⁰. In the present study, we used the Y maze test as a simple test that does not require training. In this test, behavioral variability would be relatively large because we do not provide a clear motivation such as a food reward. Therefore, the Y maze test might not be able to detect minute differences, though the Y maze test in our laboratory can detect working memory deficits by pharmacological manipulation²⁷. Therefore, we conclude that the role of dopamine D₅ receptors in working memory is limited.

Because our dopamine D5KO mice showed almost normal motor functions and working memory, we conducted the 3-CSRTT to assess impulsive actions. The q-learning analysis revealed that small deficits of learning were observed in D5KO mice (Table 1). In other words, D5KO mice, especially male mice, exhibit an inferior ability to learn from their mistakes and fine-tune their behavior. However, these small differences did not

	Wildtype littermates		D ₅ receptor KO mice	
	Male	Female	Male	Female
RNA analysis	4	4	4	4
Home cage activity	13	19	13	12
Open field test	12	17	12	12
Y maze test	12	19	13	12
Assessment of learning process in 3-choice serial reaction time task	14	11	11	10
3-choice serial reaction time task Effects of acute duloxetine injection on impulsive action	16	12	16	16

Table 2. Grouping of mice. The left column indicates the tests that mice on the right received. The numbers in each group are slightly different because we used littermates in each experiment.

significantly affect behavioral parameters in the 3-CSRTT (Fig. 4). It should be also noted that the variability in the results of each individual mouse and each session is quite high. At a minimum, we raise the possibility that the q-learning model is useful for the analysis of learning processed in the 3-CSRTT, and the detailed time series analysis could provide a clue to clarify the function of D₅ receptors.

We replicated the dose-dependent anti-impulsive effects of duloxetine previously found in male rats⁵ using male and female mice. However, the anti-impulsive effects of duloxetine were detected not only in the wild type littermates but also in dopamine D₅ receptor KO mice. That is, D₅ receptor KO failed to prevent the anti-impulsive effects of duloxetine, indicating that our original hypothesis was incorrect. Moreover, the baseline of impulsive action (following 0 mg of duloxetine) was almost the same between D₅ receptor KO mice and wild type littermates. Based on these results, we suggest that dopamine D₅ receptors do not play an important role in impulsivity. It should be noted that other parameters were also affected by duloxetine. Accuracy, a measure of attentional function, was decreased when 3 mg/kg duloxetine was injected in both genotype and both sexes. In addition, 1 mg/kg and 3 mg/kg duloxetine administration prolonged the mean correct latency and reward latency in both genotype and both sexes. These measures represent motivation and motor function. The percentage of omissions, which represents attentional function and motivation, was not affected by duloxetine. Therefore, the prolonged latencies would reflect a decrease in motor function, indicating that higher doses of duloxetine would be inappropriate in the evaluation of anti-impulsive effects. However, we still conclude that dopamine D₅ receptors have a negligible role in impulse control because these side effects were equally observed in either genotype and the anti-impulsive effects of a low or moderate dose of duloxetine did not disappear in D5KO mice.

In light of these results, how should we interpret previous studies^{5,6} indicating that drugs suppress impulsivity by stimulating dopamine D₁-like receptors in the mPFC? There are at least two possibilities. First, dopamine D₁ receptors may be more involved in impulsivity suppression, since the involvement of D₅ receptors has been ruled out. However, since dopamine D₁ receptors are also densely expressed in the nucleus accumbens, where impulsivity is enhanced by their stimulation^{7,8}, they will not be an appropriate molecular target for anti-impulsive drugs. The second possibility is that previous studies have largely examined nonselective effects of dopamine D₁-like receptor antagonists, where SCH23390 is frequently used, although its selectivity for D₁-like receptors is not high enough to completely exclude effects on other receptors and channels²⁸. In either case, the development of a selective dopamine D₅ receptor agonist would not resolve the current problems encountered in current anti-impulsive drugs. Interestingly, a recent study showed that striatal dopamine D₅ receptors are involved in the pathophysiology of levodopa-induced dyskinesia²⁹. Therefore, dopamine D₅ receptors might play a role in pathological but not physiological situations.

Materials and methods

Animals. Adult male and female D5KO mice¹⁵ or wildtype littermates (8–28 weeks old) were used. The B6.129-Drd5 <tm1Mok> mouse strain (RBRC01084) was provided by RIKEN BRC through the National Bio-Resource Project of the MEXT, Japan. In the D5KO mice used in this study, the entire dopamine D₅ receptor gene region was removed and replaced with a neomycin resistance gene. These mice were backcrossed to the C57BL/6N strain for more than 13 generations. C57BL/6N mice were supplied from Nippon SLC Co. Ltd (Hamamatsu, Japan). Animals were group-housed before starting behavioral experiments at 25 °C ± 2 °C and relative humidity of 40%–50%. Food and water were provided ad libitum except for the mice undergoing the 3-choice serial reaction time task. The lights of the animal rooms were turned on from 19:00 to 07:00. All tests were performed during the dark period except for the home cage activity test. All procedures followed the guidelines for the Care and Use of Laboratory Animals from the Animal Research Committee of Hokkaido University and were approved by the Animal Research Committee of Hokkaido University (approval no. 18-0070). We conducted all experiments in compliance with the Animal Research: Reporting of In Vivo Experiments (ARRIVE) guidelines. Mice received one or several behavioral tests as summarized in Table 2. A few mice that experienced the 3-choice serial reaction time task were excluded from assessment of learning ability because a programming error affected the premature response latency data.

Drugs. Duloxetine hydrochloride (Tokyo Chemical Industry Co., Ltd., Tokyo, Japan) was dissolved in saline and administered intraperitoneally at a volume of 10 mL/kg. Doses reported here are based on the molecular weight of the salt.

RNA analysis. Mice were deeply anesthetized with urethane (2 g/kg) intraperitoneally and sacrificed by decapitation. Brain tissue including hippocampus, medial prefrontal cortex (mPFC), and striatum (Str) were dissected on ice. Each sample was weighed, placed in a tube, immediately frozen in liquid nitrogen and kept frozen at -80°C until analysis. Total RNA was extracted from tissue using NucleoSpin RNA reagent (Takara Bio, Shiga, Japan). The mRNA expression levels of *Drd1* and *Drd5* were quantified by reverse-transcription quantitative PCR (RT-qPCR) using the respective cDNA fragment as a standard and were normalized to mouse *Gapdh* mRNA levels. Briefly, 5 μg of total RNA were reverse transcribed using ReverTra Ace[®] qPCR RT Master Mix with gDNA Remover (Toyobo, Osaka, Japan). Real-time quantitative PCR was performed on a fluorescence thermal cycler Step One[™] Real-time PCR System (Thermo Fisher Scientific, Waltham, MA, USA) by using TaqMan[®] Fast Advanced Master Mix + probe set (Thermo Fisher Scientific). The PCR conditions were 50°C for 2 min, 95°C for 20 s, followed by 40 cycles of 95°C for 1 s, and 60°C for 20 s. Primer sequences for *Drd1* (Thermo Fisher Scientific, Mm01353211_m1) and *Drd5* (Thermo Fisher Scientific, Mm00658653_s1) were chosen based on a previous study³⁰. *Gapdh* was used as a control (Thermo Fisher Scientific, Mm99999915_g1). The results were analyzed using the StepOne Software ver.2.3 (Thermo Fisher Scientific).

Home cage activity. Animals were individually housed in a Plexiglas cage (18 cm \times 26 cm \times 12 cm) for at least 1 week before this test. Spontaneous movements were measured by a passive infrared sensor that detected changes in animal thermal radiation due to movement³¹. The sensor detected a change in the intensity of infrared energy radiated from an animal (The Chronobiology Kit, Stanford Software Systems, Stanford, CA). The amount of movement was recorded every minute with computer software Analysis98 (Stanford Software Systems, Santa Cruz, CA).

Open field test. A mouse was placed in an acrylic box (45 \times 45 \times 45 cm) for 70 min. The inside of the box was covered by rough-surfaced polypropylene sheets. The light intensity in the box was adjusted to 20 lx. The movement of each mouse was monitored through a CCD camera and was tracked using a software package (LimeLight, Actimetrics, USA). We considered the total distance traveled and the number of total crossings (defined by crossings of the lines made by the division of the chamber into 7.5 cm \times 7.5 cm squares) as measures of locomotor activity. Moreover, we considered the percentage of time spent in the central area (15 cm \times 15 cm square) as a measure of anxiety-like behavior.

Y maze test. The details of the Y maze test have been described in our previous studies^{27,32}. Briefly, a mouse was placed in an apparatus consisting of three arms (10 cm-wide, 45 cm-length, and 35 cm-high-walls) for 8 min. The light intensity in the apparatus was adjusted to 20 lx. The number of entries into an arm was as a measure of locomotor activity. The percentage of spontaneous alternation was used as a measure of working memory.

3-choice serial reaction time task (3-CSRTT). Mice were trained to perform the 3-CSRTT as described previously³³. We purchased aluminum operant chambers from Med Associates Inc. (St. Albans, VT, USA). The main sequence of the 3-CSRTT is briefly described below. When a mouse entered the food magazine, a 5-s inter-trial interval (ITI) began. After the ITI, one of the three hole lights was turned on (stimulus duration (SD) in experimental sessions: 1 s (SD1)) with a pseudo-random order. Nose poking before turning on a hole light was recorded as a “premature response,” which is a measure of impulsive action. Nose poking into the lit hole was recorded as a correct response and resulted in delivery of a palatable food pellet (20 mg, dustless precision pellets, Bio-Serv, Frenchtown, NJ, USA). Nose poking into an unlit hole was recorded as an incorrect response. When the animal did not nose poke into any holes, we recorded it as an omission. A 5-s time-out period started after premature responses, incorrect responses, and omissions. We also recorded the premature response latency (the time between the ITI onset and a nose poke into a unlit hole), the correct response latency (the time between stimulus onset and a nose poke into the lit hole), and reward latency (the time between reward delivery and a nose poke into the food magazine).

Session data in the 3-CSRTT were used for two purposes (Fig. 6a). Training sessions after a pre-training period were used for q-learning analysis to assess learning ability. The pre-training sessions included several types of training and mice usually experienced each step for only a few sessions. After five SD1-ITI9 sessions were completed, duloxetine administration was started as described later.

Assessment of learning ability with a q-learning model with 3-CSRTT training sessions. We focused on ITI with 5-s (ITI5) training session data from the first session after the pre-training process to the tenth session (Fig. 6a). In these sessions, our preliminary analysis showed no clear difference in proportions of result categories (correct, premature, incorrect, or omission) between genotypes (see Supplementary Fig. 2). However, an impulsive action could be related to previous trial behaviors, and detailed analysis with a q-learning model could reveal differences between wild type and D5KO mice in terms of impulsivity. Premature response latency was recorded for these sessions and combined with correct and incorrect latencies. We could reconstruct the time between stimulus onset and a nose poke into the hole regardless of trial results.

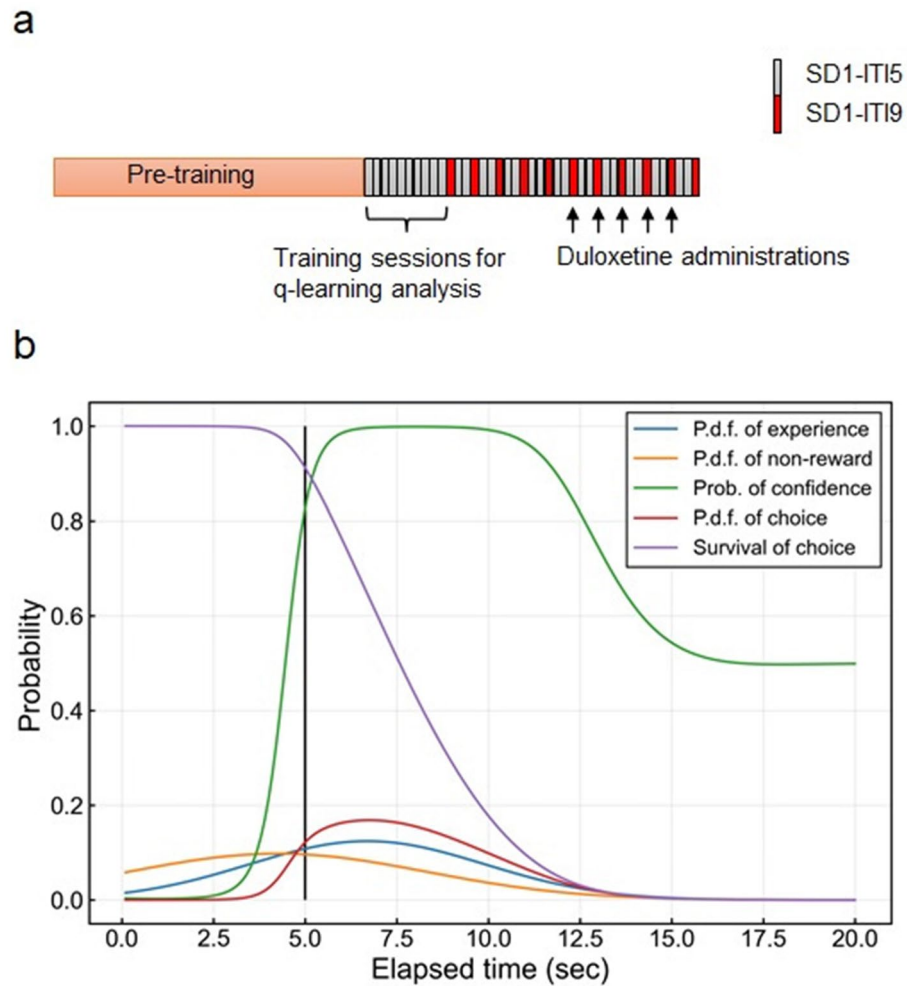


Figure 6. Sessions used for q-learning analysis and duloxetine administrations and illustration of the functions used in the q-learning model in the 3-choice serial reaction time task. **(a)** Training sessions after a pre-training period were used for q-learning analysis to assess the learning process. A gray or red box indicates one session. Duloxetine was administered after five SD1-ITI9 were completed. SD1-ITI5 stands for stimulus duration of one second and inter trial interval (ITI) of 5 s. SD1-ITI9 stands for stimulus duration of one second and inter trial interval (ITI) of 9 s. **(b)** X-axis represents elapsed time (sec) from the starting time of trials. The vertical black line at 5 s indicates the timing of the light stimulus. Each line represents the following: Probability density function (p.d.f.) of experience (blue); the p.d.f. of experience distribution. P.d.f. of non-reward (orange); the p.d.f. of non-reward distribution. Prob. of confidence (green); the probability of confidence at each elapsed time. P.d.f. of choice (red); the p.d.f. of choice distribution. Survival of choice (purple); the survival function of the choice distribution.

The q-learning model in this study attempts to capture the mechanisms of premature behavior, which represents an impulsive action. To model the memory state in the mouse brain, we assumed two types of time-dependent probability distribution functions (p.d.f.) to represent “experience” and “non-reward” memory states. The experience distribution represents the memory of all trials that mice have completed, and a non-reward distribution represents the memory of trials with premature results. Combining these two mechanisms, mice decide when to nose poke into a hole. Parameters of both distributions are updated based on the q-learning process. Since incorrect results were thought to be caused by different mechanisms and the number of incorrect results was small, incorrect results were treated the same as correct ones in this analysis.

We firstly define random variables for “experience” and “non-reward” distributions, which follow a normal distribution. With q-learning theory, these two random variables were updated based on a result type and elapsed time from a start timing of the previous trial. Rates of updating each parameter are controlled by learning rates of the experience distribution, α_X , and the non-reward distribution, α_Y . If these learning rates are lower for D5KO mice than wild-type mice, we state that D5KO mice have a deficit in learning ability.

Experience and non-reward distributions are used to calculate the probability of confidence with a softmax function. An inverse temperature, β , in the softmax function controls the degree of confidence by weighting experience and non-reward distribution. If the inverse temperature is higher for D5KO mice than wild-type

mice, D5KO mice have strong confidence in their own choices. This probability of confidence is multiplied by the experience distribution and scaled to one, yielding the probability density function of choice representing the time of the decision to nose poke into a hole. This function can be converted to be a survival function, which is used for simulation purposes. These probabilities and distributions are illustrated in Fig. 6b.

Detailed explanations of model derivation, parameter specification, estimation procedures, and simulation procedures are in Supplementary Information.

The effects of acute duloxetine injection on impulsive action in mice. To determine whether dopamine D₃ receptors play an important role in the enhancement of impulse control, we administered duloxetine (0, 0.3, 1.0, and 3.0 mg/kg) intraperitoneally to D5KO mice and their wildtype littermates 30 min before the 3-choice serial reaction time task session. We did not use higher doses of duloxetine (> 3.0 mg/kg) because higher doses induced sedation in our preliminary study. Drug treatments were carried out using a Latin square design and were administered with at least a 2-day interval between injections. During the testing phase of this study, the duration of the ITI was prolonged to 9 s (ITI9) because the mice made only a few (< 10) premature responses during the task using a 5-s interval (ITI5). Each testing session with ITI9 was conducted for 70 min or until 100 trials were completed, whichever came first, while sessions with ITI5 were conducted for 60 min or until 100 trials were completed, whichever came first. When the mice experienced 10 ITI5 sessions, they were habituated to ITI9 sessions 6 times with 2-day intervals.

The following behavioral measures in the 3-CSRTT were analyzed:

- Percentage of premature responses: $[\text{premature responses}/(\text{premature} + \text{correct} + \text{incorrect responses})] \times 100$, a measure of impulsive action
- Accuracy (percentage of correct responses): $[\text{correct responses}/(\text{correct} + \text{incorrect responses})] \times 100$, a measure of attentional function
- Percentage of omissions $[(\text{number of omissions}/\text{total initiated trials}) \times 100]$, a measure of attentional function and motivation for the task
- Correct response latency (s), a measure of attentional function, motivation for the task, and motor function
- Reward latency (s), a measure of motivation for reward and motor function

Statistical analysis. For the measurement of locomotor activity in the home cage and in the open field test, we used a three-factor mixed analysis of variance (ANOVA) with time as a within-subjects factor and genotype and sex as between-subjects factors. For the effect of genotype in the Y maze test, we used a two-factor mixed ANOVA with genotype and sex as between-subjects factors. For the 3-CSRTT, each measure was analyzed separately by a three-factor mixed ANOVA with drug as a within-subjects factor and genotype and sex as between-subjects factors except for the assessment of learning ability. If Mauchly's sphericity test was significant, a Greenhouse–Geisser correction was used. Multiple comparisons with Bonferroni's correction were also conducted in cases where ANOVA revealed a significant main effect. All results except for the assessment of learning ability are presented as mean \pm standard error of the mean (S.E.M.). The results were considered statistically significant when $p < 0.05$. SPSS (version 23.0) and GraphPad Prism (version 8.4.2) were used for statistical analyses.

Data availability

The datasets of this study are available from the corresponding author on reasonable request.

Code availability

Codes used for model fitting and plotting of q-learning assessments of the 3-choice serial reaction time task is available on a GitHub repository at https://github.com/Neuropharmacol/3csrtt_q_learning.

Received: 9 December 2021; Accepted: 7 March 2022

Published online: 10 April 2022

References

- Moeller, F. G., Barratt, E. S., Dougherty, D. M., Schmitz, J. M. & Swann, A. C. Psychiatric aspects of impulsivity. *Am. J. Psychiatry* **158**, 1783–1793. <https://doi.org/10.1176/appi.ajp.158.11.1783> (2001).
- Chen, Q. *et al.* Common psychiatric and metabolic comorbidity of adult attention-deficit/hyperactivity disorder: A population-based cross-sectional study. *PLoS ONE* **13**, e0204516. <https://doi.org/10.1371/journal.pone.0204516> (2018).
- Ayerbe, L. *et al.* Hypertension risk and clinical care in patients with bipolar disorder or schizophrenia; a systematic review and meta-analysis. *J. Affect. Disord.* **225**, 665–670. <https://doi.org/10.1016/j.jad.2017.09.002> (2018).
- El-Gabalawy, R., Katz, L. Y. & Sareen, J. Comorbidity and associated severity of borderline personality disorder and physical health conditions in a nationally representative sample. *Psychosom. Med.* **72**, 641–647. <https://doi.org/10.1097/PSY.0b013e3181e10c7b> (2010).
- Sasamori, H., Ohmura, Y., Yoshida, T. & Yoshioka, M. Noradrenergic reuptake inhibition increases control of impulsive action by activating D1-like receptors in the infralimbic cortex. *Eur. J. Pharmacol.* **844**, 17–25. <https://doi.org/10.1016/j.ejphar.2018.11.041> (2019).
- Tsutsui-Kimura, I. *et al.* Milnacipran enhances the control of impulsive action by activating D(1)-like receptors in the infralimbic cortex. *Psychopharmacology* **225**, 495–504. <https://doi.org/10.1007/s00213-012-2835-5> (2013).
- Cole, B. J. & Robbins, T. W. Effects of 6-hydroxydopamine lesions of the nucleus accumbens septi on performance of a 5-choice serial reaction time task in rats: Implications for theories of selective attention and arousal. *Behav. Brain Res.* **33**, 165–179 (1989).
- Pattij, T., Janssen, M. C., Vanderschuren, L. J., Schoffelmeier, A. N. & van Gaalen, M. M. Involvement of dopamine D1 and D2 receptors in the nucleus accumbens core and shell in inhibitory response control. *Psychopharmacology* **191**, 587–598. <https://doi.org/10.1007/s00213-006-0533-x> (2007).

9. Sarinana, J. & Tonegawa, S. Differentiation of forebrain and hippocampal dopamine 1-class receptors, D1R and D5R, in spatial learning and memory. *Hippocampus* **26**, 76–86. <https://doi.org/10.1002/hipo.22492> (2016).
10. Sokolowski, J. D. & Salamone, J. D. Effects of dopamine depletions in the medial prefrontal cortex on DRL performance and motor activity in the rat. *Brain Res.* **642**, 20–28 (1994).
11. Tsutsui-Kimura, I., Ohmura, Y., Yoshida, T. & Yoshioka, M. Milnacipran affects mouse impulsive, aggressive, and depressive-like behaviors in a distinct dose-dependent manner. *J. Pharmacol. Sci.* <https://doi.org/10.1016/j.jphs.2017.06.004> (2017).
12. Sunahara, R. K. *et al.* Cloning of the gene for a human dopamine D5 receptor with higher affinity for dopamine than D1. *Nature* **350**, 614–619. <https://doi.org/10.1038/350614a0> (1991).
13. Gardner, E. L. Addiction and brain reward and antireward pathways. *Adv. Psychosom. Med.* **30**, 22–60. <https://doi.org/10.1159/000324065> (2011).
14. Hollon, T. R. *et al.* Mice lacking D5 dopamine receptors have increased sympathetic tone and are hypertensive. *J. Neurosci.* **22**, 10801–10810. <https://doi.org/10.1523/jneurosci.22-24-10801.2002> (2002).
15. Hayashizaki, S. *et al.* Methamphetamine increases locomotion and dopamine transporter activity in dopamine d5 receptor-deficient mice. *PLoS ONE* **8**, e75975. <https://doi.org/10.1371/journal.pone.0075975> (2013).
16. El-Brolosy, M. A. *et al.* Genetic compensation triggered by mutant mRNA degradation. *Nature* **568**, 193–197. <https://doi.org/10.1038/s41586-019-1064-z> (2019).
17. Ma, Z. *et al.* PTC-bearing mRNA elicits a genetic compensation response via Upf3a and COMPASS components. *Nature* **568**, 259–263. <https://doi.org/10.1038/s41586-019-1057-y> (2019).
18. Nakamoto, C. *et al.* GluD1 knockout mice with a pure C57BL/6N background show impaired fear memory, social interaction, and enhanced depressive-like behavior. *PLoS ONE* **15**, e0229288. <https://doi.org/10.1371/journal.pone.0229288> (2020).
19. Crawley, J. N. *et al.* Behavioral phenotypes of inbred mouse strains: Implications and recommendations for molecular studies. *Psychopharmacology* **132**, 107–124. <https://doi.org/10.1007/s002130050327> (1997).
20. Carr, G. V., Maltese, F., Sibley, D. R., Weinberger, D. R. & Papaleo, F. The dopamine D5 receptor is involved in working memory. *Front. Pharmacol.* **8**, 666. <https://doi.org/10.3389/fphar.2017.00666> (2017).
21. Holmes, A. *et al.* Behavioral characterization of dopamine D5 receptor null mutant mice. *Behav. Neurosci.* **115**, 1129–1144 (2001).
22. Moraga-Amaro, R. *et al.* Dopamine receptor D5 deficiency results in a selective reduction of hippocampal NMDA receptor subunit NR2B expression and impaired memory. *Neuropharmacology* **103**, 222–235. <https://doi.org/10.1016/j.neuropharm.2015.12.018> (2016).
23. Tsutsui-Kimura, I. *et al.* The effects of serotonin and/or noradrenergic reuptake inhibitors on impulsive-like action assessed by the three-choice serial reaction time task: A simple and valid model of impulsive action using rats. *Behav. Pharmacol.* **20**, 474–483. <https://doi.org/10.1097/FBP.0b013e3283305e65> (2009).
24. Laplante, F., Sibley, D. R. & Quirion, R. Reduction in acetylcholine release in the hippocampus of dopamine D5 receptor-deficient mice. *Neuropsychopharmacology* **29**, 1620–1627. <https://doi.org/10.1038/sj.npp.1300467> (2004).
25. Perreault, M. L., Jones-Tabah, J., O'Dowd, B. F. & George, S. R. A physiological role for the dopamine D5 receptor as a regulator of BDNF and Akt signalling in rodent prefrontal cortex. *Int. J. Neuropsychopharmacol.* **16**, 477–483. <https://doi.org/10.1017/s1461145712000685> (2013).
26. Sahu, A., Tyeryar, K. R., Vongtau, H. O., Sibley, D. R. & Undieh, A. S. D5 dopamine receptors are required for dopaminergic activation of phospholipase C. *Mol. Pharmacol.* **75**, 447–453. <https://doi.org/10.1124/mol.108.053017> (2009).
27. Ohmura, Y. *et al.* Disruption of model-based decision making by silencing of serotonin neurons in the dorsal raphe nucleus. *Curr. Biol.* <https://doi.org/10.1016/j.cub.2021.03.048> (2021).
28. Kuzhikandathil, E. V. & Oxford, G. S. Classic D1 dopamine receptor antagonist R-(+)-7-chloro-8-hydroxy-3-methyl-1-phenyl-2,3,4,5-tetrahydro-1H-3-benzazepine hydrochloride (SCH23390) directly inhibits G protein-coupled inwardly rectifying potassium channels. *Mol. Pharmacol.* **62**, 119–126. <https://doi.org/10.1124/mol.62.1.119> (2002).
29. Wang, Y. *et al.* Inhibition of striatal dopamine D(5) receptor attenuates levodopa-induced dyskinesia in a rat model of Parkinson's disease. *Brain Res.* **1754**, 147266. <https://doi.org/10.1016/j.brainres.2020.147266> (2021).
30. McCarthy, D. M. *et al.* Nicotine exposure of male mice produces behavioral impairment in multiple generations of descendants. *PLoS Biol.* **16**, e2006497. <https://doi.org/10.1371/journal.pbio.2006497> (2018).
31. Abe, H., Honma, S., Ohtsu, H. & Honma, K. Circadian rhythms in behavior and clock gene expressions in the brain of mice lacking histidine decarboxylase. *Brain Res. Mol. Brain Res.* **124**, 178–187. <https://doi.org/10.1016/j.molbrainres.2004.02.015> (2004).
32. Hamadate, N. *et al.* Liposome-encapsulated hemoglobin ameliorates impairment of fear memory and hippocampal dysfunction after cerebral ischemia in rats. *J. Pharmacol. Sci.* **114**, 409–419. <https://doi.org/10.1254/jphs.10207fp> (2010).
33. Sasamori, H., Ohmura, Y., Kubo, T., Yoshida, T. & Yoshioka, M. Assessment of impulsivity in adolescent mice: A new training procedure for a 3-choice serial reaction time task. *Behav. Brain Res.* **343**, 61–70. <https://doi.org/10.1016/j.bbr.2018.01.014> (2018).

Acknowledgements

We would like to thank Dr. Motoya Katsuki for providing dopamine D₅ receptor KO mice. We would like to thank Tomoko Furukawa, Aki Tanimori, and Dr. Mao Nebuka for their help in breeding the transgenic mice and supporting behavioral experiments, and thank JAM Post (<https://www.jamp.com/index.cfm>) for the English language review.

Author contributions

H.S. contributed to the 3-CSRTT experiments, study design, interpretation, analyses, and manuscript preparation. T.A. contributed to the study design, interpretation, analyses using a q-learning model, and manuscript preparation. C.S. contributed to all of the experiments except for the 3-CSRTT. Y.B., N.N., M.S., and T.Y. contributed to the study design and manuscript revision. Miwako Yamasaki and M.W. contributed to providing D5KO mice and interpretation of the results. A.T. contributed to the real-time RT-PCR study design and experiments. Y.O. contributed to the conception and the study design, interpretation, analyses, and manuscript preparation. Mitsuhiro Yoshioka contributed to the study design and interpretation, but he passed away before completing this project. All authors, except for Mitsuhiro Yoshioka, reviewed the manuscript.

Funding

This work was supported by JSPS KAKENHI Grant Numbers JP21K20856 (HS), JP19J20112 (HS), 18K07545 (YO), and 19H04976 (YO).

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-022-10013-5>.

Correspondence and requests for materials should be addressed to Y.O.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022